

Forum Editorial

Redox Signaling in Cancer Biology

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ABSTRACT

Over the last three decades, it has become increasingly clear that intracellular signaling pathways are activated via changes in intracellular metabolic oxidation/reduction (redox) reactions involving reactive oxygen species (ROS; *i.e.*, superoxide and hydrogen peroxide). The initial proposals hypothesized that signaling through metabolic oxidation/reduction (redox) reactions involving ROS could contribute to carcinogenesis and progression to malignancy. Strong evidence for this hypothesis was obtained from studies showing that environmental insults (*i.e.*, ionizing radiation) as well as xenobiotics (*i.e.*, polycyclic aromatic hydrocarbons and phorbol esters) capable of inducing steady-state increases in free radical production and ROS could act as both initiators and promoters of carcinogenesis. This Forum is directed at understanding possible redox signaling mechanisms governing cellular radiation response, tumor growth, and response to therapy, as well as the role of nitric oxide in cancer biology. *Antioxid. Redox Signal.* 8, 1249–1252.

OVER THE LAST THREE DECADES, it has become increasingly clear that intracellular signaling pathways are activated via changes in intracellular metabolic oxidation/reduction (redox) reactions involving reactive oxygen species (ROS; *i.e.*, superoxide and hydrogen peroxide) (1, 2, 4, 6, 21, 22, 24, 28, 29, 32). The initial proposals hypothesized that signaling through metabolic oxidation/reduction (redox) reactions involving ROS could contribute to carcinogenesis and progression to malignancy (2, 6, 21, 22, 24). Strong evidence for this hypothesis was obtained from studies showing that environmental insults (*i.e.*, ionizing radiation) as well as xenobiotics (*i.e.*, polycyclic aromatic hydrocarbons and phorbol esters) capable of inducing steady-state increases in free radical production and ROS could act as both initiators and promoters of carcinogenesis (reviewed in 2, 6, 21, 22, 24, 28, 29). This was thought to occur by increasing mutagenesis, inhibiting differentiation, converting protooncogenes into oncogenes as well as inactivating tumor-suppressor genes, and stimulation of mitogenesis (2, 6, 21, 22, 24, 25, 28, 29, 32).

Further support for a prooxidant environment in cancer cells came from studies showing that in general cancer cells showed altered expression of cellular antioxidants that metabolized ROS, particularly Mn superoxide dismutase and enzyme systems involved in hydroperoxide metabolism, such as catalase (21, 22, 24). In addition, reports suggested that cancer cells produced increased steady-state levels of ROS (particularly hydroperoxides), relative to normal cells, presumably from some metabolic process (5, 29, 34). Furthermore, dietary antioxidants were shown to act as anticarcinogens in a variety of models of malignant transformation (2, 6). These results all provided strong support for the hypothesis that increases in the steady-state levels of free radicals and ROS in mammalian cells could lead to a prooxidant state that was causally involved with malignant transformation (2, 6, 21, 22, 24, 28, 29).

As the body of evidence supporting the involvement of redox reactions in the malignant transformation process was accumulating, a clearer mechanistic understanding of the involvement of redox signaling in normal cellular processes

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emerged. In general, protein kinases were found to be activated and protein phosphatases were found to be inhibited by exposure to oxidants (7, 13, 30, 33, 36). A multitude of transcription factors were discovered to contain redox regulatory sites (primarily cysteine residues), and mutations in these regulatory sites were shown to convert protooncogenes into oncogenes as well as inactivating tumor-suppressor genes (1, 25, 32). Furthermore, cell growth signaling was found to be stimulated by oxidants (33), and growth factors were discovered to stimulate cell division by activating redox-sensitive ROS-mediated signaling pathways thought to involve oxidase enzymes (*i.e.*, NADPH-oxidases) as well as mitochondrial metabolism (4, 10, 18, 19, 33). Finally, increasing steady-state levels of metabolic ROS production by overexpressing NADPH-oxidase enzymes or mutations in genes coding for mitochondrial electron transport chain proteins was shown to increase transformation (14, 31). These findings all support the hypothesis that normal cell-signaling processes involve many redox-sensitive regulatory sites and that these signaling processes can be disrupted by changes in oxidative metabolism that are associated with neoplastic transformation. This has led to the suggestion that cancer could represent a constellation of metabolic and/or genetic diseases in which the common theme is uncoupling of normal cellular processes that govern cell growth and development caused by the inappropriate flow of electrons from metabolic oxidation/reduction reactions to redox-sensitive signal-transduction and gene-expression pathways (29).

In addition to the aforementioned evidence that redox signaling was involved with the process of transformation, evidence emerged that altered metabolic redox states in cancer cells could be exploited to kill cancer cells selectively, relative to normal cells. Initially it was shown that the toxicity of chemotherapeutic agents as well as radiation toward cancer cells could be enhanced by agents that inhibited the function of thiol-dependent (*i.e.*, glutathione and thioredoxin) antioxidant detoxification systems involved with resistance to therapy (20, 39). It was also hypothesized that treating patients with exogenous thiol antioxidants could selectively protect normal tissues from oxidative damage during chemotherapy and radiation therapy (11, 37, 38) and improve outcome. More recently, it has been proposed that treating cancer cells with vectors causing the overexpression of MnSOD and catalase could inhibit tumor cell growth, presumably by disrupting redox signaling involved with cell division (23) as well as protecting normal tissues from therapeutic intervention (12). Finally, it has been shown that cancer cells are more susceptible than normal cells to glucose deprivation-induced metabolic oxidative stress (5, 29). This was hypothesized to be the result of increased glucose metabolism in cancer cells to obtain reducing equivalents to compensate for increased steady-state levels of hydroperoxide production by defective mitochondrial oxidative metabolism (5, 29). Furthermore, inhibitors of glucose and hydroperoxide metabolism have been suggested to enhance metabolic oxidative stress selectively in cancer cells (relative to normal cells), leading to both radio- and chemosensitization (3, 16). Taken together, the current evidence presented in this overview demonstrates that disruptions in metabolic redox signaling contribute to both the genesis and maintenance of the malignant phenotype, as well as

potentially providing a biochemical rationale for selective killing of cancer cells, relative to normal cells. With this background in mind, the following contributions were chosen for this Forum Issue to highlight recent developments in the field.

The original research contributions fall into two main categories relevant to redox signaling and cancer biology. The first group is directed at understanding possible mechanisms governing cellular radiation response. In Pandey *et al.* (26), the effects of low- and high-dose radiation on mitochondrial membrane potential and protein import were found to be quite different; with low-dose radiation stimulating and high-dose radiation inhibiting protein import and membrane potential in normal human fibroblasts. The authors suggest that these differential effects on mitochondrial metabolic state and subsequent redox signaling may have significant physiologic implications after high and low doses of radiation. Cook *et al.* (8) studied the baseline and radiation-induced gene-expression profiles of murine head and neck cancer cells exposed *in vitro* and *in vivo* and found that the gene-expression profiles from the same tumor cells grown under different metabolic conditions differed significantly, as did the expression changes induced by radiation exposure. They also found that the expression of a significant number of genes governing metabolic redox responses were among those affected by ionizing radiation. These results provide further support for the hypothesis that exposure of cancer cells to ionizing radiation causes changes in gene expression and metabolic redox signaling that could contribute to radiation responses. This work also highlights the need to control initial environmental conditions carefully before making conclusions about the relative importance of gene-expression changes to biologic responses. In the final article studying radiation effects, Kalen *et al.* (15) found that prolonged stable overexpression of the mitochondrial form of the superoxide dismutase enzyme (MnSOD) in human head and neck cancer cells enhanced radiation-induced G_2 accumulation and radioresistance. Because MnSOD is an enzyme thought to modify intracellular redox signaling and cell-cycle progression, these results support the hypothesis that mitochondrially derived metabolic redox signaling can modify radiation responses by altering cell-cycle progression and the G_2 -checkpoint pathway.

The second group of original contributions is directed at elucidating redox signaling pathways that affect cancer cell responses relevant to inhibiting tumor growth and cancer therapy. By using site-directed mutagenesis of the active site, Zhang *et al.* (40) determined that the growth-suppressive effects of overexpressing MnSOD in transformed cells is caused by the enzymatic activity of the protein. These results support the idea that increasing the dismutation of mitochondrial superoxide alters redox signaling in cancer cells that can result in growth suppression. In addition, it suggests that either steady-state levels of mitochondrial superoxide may be involved in maintaining cancer cell growth or hydrogen peroxide (the product of the dismutation reaction) may be involved in inhibiting tumor cell growth. In the article by Dasgupta *et al.* (9), they show that stable overexpression of MnSOD protects human fibrosarcoma cells from tumor necrosis factor-mediated apoptosis. Based on these results, they suggest that increasing steady-state concentrations of

hydrogen peroxide, by overexpressing MnSOD, inactivate redox-sensitive signaling leading to caspase activation (9). These two articles (Zhang *et al.* and Dasgupta *et al.*) coupled with the Kalen *et al.* article also bring up the important point that MnSOD overexpression can have beneficial as well as deleterious effects on cell growth when developing cancer therapeutic strategies. One possible explanation for this apparent paradox is likely to involve the overall redox balance in the cancer cells at the time of treatment. Based on these observations, it is tempting to speculate that a key determinant of outcome in cancer cells overexpressing MnSOD may be the expression of hydrogen peroxide scavenging systems (*i.e.*, catalase, glutathione peroxidases, and/or peroxiredoxins); however, this awaits further verification.

Luo *et al.* (17) show that inhibition of MnSOD, CuZn-SOD, and catalase activities can be accomplished by treating cells with singlet oxygen derived from photodynamic therapy. These results suggest that photodynamic therapy can be used in conjunction with other anticancer agents that increase superoxide and hydrogen peroxide production to improve tumor cell killing in combined-modality cancer therapy protocols. The final article in this group by Tome *et al.* (35) shows that overexpression of the first enzyme in the pentose phosphate cycle (where NADPH is regenerated from NADP⁺) sensitizes murine thymic lymphoma cells to cell killing mediated by several standard chemotherapeutic agents. These results encourage speculation that excess reducing equivalents, in the form of NADPH, can sensitize cancer cells to redox signaling, resulting in apoptosis (possibly through NADPH-dependent oxidase enzymes) during combined-modality chemotherapy.

The last forum contribution by Ridnour *et al.* (27) reviews and highlights the exciting field of nitric oxide-mediated redox signaling (in addition to ROS-mediated signaling) and its relevance to both cytoprotective and cytotoxic responses in cancer biology. Finally, the guest editors thank all the authors for their excellent and provocative contributions to this forum on redox signaling and cancer biology. We also hope that the readers of this issue will become inspired to make scientific contributions from their own laboratories to this growing field of study.

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ABBREVIATIONS

CuZnSOD, copper zinc superoxide dismutase; MnSOD, manganese superoxide dismutase; NADPH/NADP⁺, nicotinamide adenine dinucleotide phosphate; ROS, reactive oxygen species.

REFERENCES

1. Abate C, Patel L, Rauscher FJ, and Curran T. Redox regulation of Fos and Jun DNA-binding activity in vitro. *Science* 249: 1157–1161, 1990.
2. Ames BN. Dietary carcinogens and anticarcinogens: Oxygen radicals and degenerative diseases. *Science* 221: 1256–1262, 1983.
3. Andringa KK, Coleman MC, Aykin-Burns N, Hitchler MJ, Walsh SA, Domann FE, and Spitz DR. Inhibition of glutamate cysteine ligase (GCL) activity sensitizes human breast cancer cells to the toxicity of 2-deoxy-D-glucose. *Cancer Res* 66: 1605–1610, 2006.
4. Arnold RS, Shi J, Murad E, Whalen AM, Sun CQ, Polavarapu R, Parthasarathy S, Petros JA, and Lambeth JD. Hydrogen peroxide mediates the cell growth and transformation caused by the mitogenic oxidase Nox1. *Proc Natl Acad Sci U S A* 98: 5550–5555, 2001.
5. Blackburn RV, Spitz DR, Liu X, Galoforo SS, Sim JE, Ridnour LA, Chen JC, Davis BH, Corry PM, and Lee YJ. Metabolic oxidative stress activates signal transduction and gene expression during glucose deprivation in human tumor cells. *Free Radic Biol Med* 26: 419–430, 1999.
6. Cerutti PA. Prooxidant states and tumor promotion. *Science* 227: 375–381, 1985.
7. Claiborne A, Yeh JJ, Mallett TC, Luba J, Crane EJ, Charrier V, and Parsonage D. Protein-sulfenic acids: diverse roles for an unlikely player in enzyme catalysis and redox regulation. *Biochemistry* 38: 15407–15416, 1999.
8. Cook JA, Chuang EY, Tsai M-H, Coffin D, DeGraff W, Sowers AL, and Mitchell JB. Radiation-induced changes in gene expression profiles for the SCC VII tumor cells grown in vitro and in vivo. *Antioxid Redox Signal* (this issue).
9. Dasgupta J, Subbaram S, Connor KM, Rodriguez AM, Beckman JS, Tirosh O, and Melendez JA. Manganese superoxide dismutase protects from TNF- α induced apoptosis by increasing the steady-state production of hydrogen peroxide. *Antioxid Redox Signal* (this issue).
10. Felty Q and Roy D. Mitochondrial signals to nucleus regulate estrogen-induced cell growth. *Med Hypotheses* 64: 133–141, 2005.
11. Gon Y, Sasada T, Matsui M, Hashimoto S, Takagi Y, Iwata S, Wada H, Horie T, and Yodoi J. Expression of thioredoxin in bleomycin-injured airway epithelium: possible role of protection against bleomycin induced epithelial injury. *Life Sci* 68:1877–1888, 2001.
12. Greenberger JS, Epperly MW, Gretton J, Jefferson M, Nie S, Bernarding M, Kagan V, and Guo HL. Radioprotective gene therapy. *Curr Gene Ther* 3: 183–195, 2003.
13. Guyton KZ, Liu Y, Gorospe M, Xu Q, and Holbrook NJ. Activation of mitogen-activated protein kinase by H₂O₂. *J Biol Chem* 271: 4138–4142, 1996.
14. Ishii T, Yasuda K, Akatsuka A, Hino O, Hartman PS, and Ishii N. A mutation in the SDHC gene of complex II increases oxidative stress, resulting in apoptosis and tumorigenesis. *Cancer Res* 65: 203–209, 2005.
15. Kalen AL, Sarsour EH, Venkataraman S, and Goswami PC. Mn-superoxide dismutase over expression enhances G₂-accumulation and radioresistance in human oral squamous carcinoma cells. *Antioxid Redox Signal* (this issue).

16. Lin X, Zhang F, Bradbury CW, Kaushal A, Li L, Spitz DR, Aft R, and Gius D. 2-Deoxy-D-glucose-induced cytotoxicity and radiosensitization in tumor cells is mediated via disruptions in thiol metabolism. *Cancer Res* 63:3413–3417, 2003.
17. Luo J, Li L, Zhang Y, Spitz DR, Buettner GR, Oberley LW, and Domann FE. Inactivation of primary antioxidant enzymes in mouse keratinocytes by photodynamically generated singlet oxygen. *Antioxid Redox Signal* (this issue).
18. Menon SG, Coleman MC, Walsh SA, Spitz DR, and Goswami PC. Differential susceptibility of nonmalignant human breast epithelial cells and breast cancer cells to thiol antioxidant-induced G₁-delay. *Antioxid Redox Signal* 7: 711–8, 2005.
19. Menon SG, Sarsour EH, Spitz DR, Higashikubo R, Sturm M, Zhang H, and Goswami PC. Redox regulation of the G₁ to S phase transition in the mouse embryo fibroblast cell cycle. *Cancer Res* 63: 2109–2117, 2003.
20. Mitchell JB and Russo A. The role of glutathione in radiation and drug induced cytotoxicity. *Br J Cancer Suppl* 8: 96–104, 1987.
21. Oberley LW and Buettner GR. Role of superoxide dismutase in cancer: a review. *Cancer Res* 39:1141–1149, 1979.
22. Oberley LW, Oberley TD, and Buettner GR. Cell division in normal and transformed cells: the possible role of superoxide and hydrogen peroxide. *Med Hypotheses* 7:21–42, 1981.
23. Oberley LW. Anticancer therapy by overexpression of superoxide dismutase. *Antioxid Redox Signal* 3:461–472, 2001.
24. Oberley LW. Superoxide dismutase and cancer. In: Oberley LW (ed). *Superoxide Dismutase*, Vol II. Boca Raton, FL: CRC Press, pp 127–166, 1982.
25. Okuno H, Akahori A, Sato H, Xanthoudakis S, Curran T, and Iba H. Escape from redox regulation enhances the transforming activity of Fos. *Oncogene* 8: 695–701, 1993.
26. Pandey BN, Gordon DM, de Toledo SM, Pain D, and Azzam EI. Normal human fibroblasts exposed to high or low dose ionizing radiation: differential effects on mitochondrial protein import and membrane potential. *Antioxid Redox Signal* (this issue).
27. Ridnour LA, Thomas DD, Donzelli S, Espey MG, Roberts DD, Wink DA, and Isenberg JS. The biphasic nature of nitric oxide responses in tumor biology. *Antioxid Redox Signal* (this issue).
28. Spitz DR, Azzam EI, Li JJ, and Gius D. Metabolic oxidation/reduction reactions and cellular responses to ionizing radiation: a unifying concept in stress response biology. *Cancer Metastasis Rev* 23: 311–322, 2004.
29. Spitz DR, Sim JE, Ridnour LA, Galoforo SS, and Lee YJ. Glucose deprivation-induced oxidative stress in human tumor cells: a fundamental defect in metabolism? *Ann NY Acad Sci* 899: 349–362, 2000.
30. Stevenson MA, Pollock SS, Coleman CN, and Calderwood SK. X-irradiation, phorbol esters, and H₂O₂ stimulate mitogen-activated protein kinase activity in NIH-3T3 cells through the formation of reactive oxygen intermediates. *Cancer Res* 54:12–15, 1994.
31. Suh YA, Arnold RS, Lassegue B, Shi J, Xu X, Sorescu D, Chung AB, Griendling KK, and Lambeth JD. Cell transformation by the superoxide-generating oxidase Mox1. *Nature* 401: 79–82, 1999.
32. Sun Y and Oberley LW. Redox regulation of transcriptional activators. *Free Radic Biol Med* 21: 335–348, 1996.
33. Sundaresan M, Yu ZX, Ferrans VJ, Irani K, and Finkel T. Requirement for generation of H₂O₂ for platelet-derived growth factor signal transduction. *Science* 270: 296–299, 1995.
34. Szatrowski TP and Nathan CF. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res* 51: 794–798, 1991.
35. Tome ME, Johnson DBF, Samulitis B, Dorr RT, and Briehl MM. Glucose 6-phosphate dehydrogenase overexpression models glucose deprivation and sensitizes lymphoma cells to apoptosis. *Antioxid Redox Signal* (this issue).
36. Tonks NK. Redox redux: revisiting PTPs and the control of cell signaling. *Cell* 121: 667–670, 2005.
37. Watson WH, Yang X, Choi YE, Jones DP, and Kehrer JP. Thioredoxin and its role in toxicology. *Toxicol Sci* 78: 3–14, 2004.
38. Weiss JF and Landauer MR. Protection against ionizing radiation by antioxidant nutrients and phytochemicals. *Toxicology* 189: 1–20, 2003.
39. Yokomizo A, Ono M, Nanri H, Makino Y, Ohga T, Wada M, Okamoto T, Yodoi J, Kuwano M, and Kohno K. Cellular levels of thioredoxin associated with drug sensitivity to cisplatin, mitomycin C, doxorubicin, and etoposide. *Cancer Res* 55: 4293–4296, 1995.
40. Zhang Y, Smith BJ, and Oberley LW. Enzymatic activity is necessary for the tumor suppressive effects of MnSOD. *Antioxid Redox Signal* (this issue).

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2. Michael J. Hitchler, Frederick E. Domann. 2012. Redox regulation of the epigenetic landscape in Cancer: A role for metabolic reprogramming in remodeling the epigenome. *Free Radical Biology and Medicine* . [[CrossRef](#)]
3. Yueming Zhu, Seong-Hoon Park, Ozkan Ozden, Hyun-Seok Kim, Haiyan Jiang, Athanassios Vassilopoulos, Douglas R. Spitz, David Gius. 2012. Exploring the electrostatic repulsion model in the role of Sirt3 in directing MnSOD acetylation status and enzymatic activity. *Free Radical Biology and Medicine* **53**:4, 828-833. [[CrossRef](#)]
4. Aaron K. Holley, Sanjit Kumar Dhar, Daret K. St. Clair. 2012. Curbing cancer's sweet tooth: Is there a role for MnSOD in regulation of the Warburg effect?. *Mitochondrion* . [[CrossRef](#)]
5. Maung Kyaw Khaing Oo, Yamin Yang, Yue Hu, Maria Gomez, Henry Du, Hongjun Wang. 2012. Gold Nanoparticle-Enhanced and Size-Dependent Generation of Reactive Oxygen Species from Protoporphyrin IX. *ACS Nano* 120309142006005. [[CrossRef](#)]
6. Apollina Goel, Douglas R. Spitz, George J. Weiner. 2012. Manipulation of cellular redox parameters for improving therapeutic responses in B-cell lymphoma and multiple myeloma. *Journal of Cellular Biochemistry* **113**:2, 419-425. [[CrossRef](#)]
7. Lyndsay M. Randolph, Miao-Ping Chien, Nathan C. Gianneschi. 2012. Biological stimuli and biomolecules in the assembly and manipulation of nanoscale polymeric particles. *Chemical Science* . [[CrossRef](#)]
8. Zhivko Zhelev, Veselina Gadjeva, Ichio Aoki, Rumiana Bakalova, Tsuneo Saga. 2012. Cell-penetrating nitroxides as molecular sensors for imaging of cancer in vivo, based on tissue redox activity. *Molecular BioSystems* **8**:10, 2733. [[CrossRef](#)]
9. Yuxing Zhang , Yanzhi Du , Weidong Le , Kankan Wang , Nelly Kieffer , Ji Zhang . 2011. Redox Control of the Survival of Healthy and Diseased Cells. *Antioxidants & Redox Signaling* **15**:11, 2867-2908. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
10. Andrei V. Budanov . 2011. Stress-Responsive Sestrins Link p53 with Redox Regulation and Mammalian Target of Rapamycin Signaling. *Antioxidants & Redox Signaling* **15**:6, 1679-1690. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
11. Hongliang Li, Yong Wang, Senthil K. Pazhanisamy, Lijian Shao, Ines Batinic-Haberle, Aimin Meng, Daohong Zhou. 2011. Mn(III) meso-tetrakis-(N-ethylpyridinium-2-yl) porphyrin mitigates total body irradiation-induced long-term bone marrow suppression. *Free Radical Biology and Medicine* **51**:1, 30-37. [[CrossRef](#)]
12. Francis Thomas, Jeff M.P. Holly, Rajendra Persad, Amit Bahl, Claire M. Perks. 2011. Green Tea Extract (Epigallocatechin-3-Gallate) Reduces Efficacy of Radiotherapy on Prostate Cancer Cells. *Urology* . [[CrossRef](#)]
13. José L. Sardina, Guillermo López-Ruano, Beatriz Sánchez-Sánchez, Marcial Llanillo, Angel Hernández-Hernández. 2011. Reactive oxygen species: Are they important for haematopoiesis?. *Critical Reviews in Oncology/Hematology* . [[CrossRef](#)]
14. Aram You, Chang-won Nam, Nobunao Wakabayashi, Masayuki Yamamoto, Thomas W. Kensler, Mi-Kyoung Kwak. 2011. Transcription factor Nrf2 maintains the basal expression of Mdm2: An implication of the regulation of p53 signaling by Nrf2. *Archives of Biochemistry and Biophysics* **507**:2, 356-364. [[CrossRef](#)]
15. Adam J. Case, Jodi L. McGill, Lorraine T. Tygrett, Takuji Shirasawa, Douglas R. Spitz, Thomas J. Waldschmidt, Kevin L. Legge, Frederick E. Domann. 2011. Elevated mitochondrial superoxide disrupts normal T cell development, impairing adaptive immune responses to an influenza challenge. *Free Radical Biology and Medicine* **50**:3, 448-458. [[CrossRef](#)]
16. Kenneth D Tew, Danyelle M Townsend. 2011. Redox platforms in cancer drug discovery and development. *Current Opinion in Chemical Biology* **15**:1, 156-161. [[CrossRef](#)]
17. Jung-Hoon Kang. 2010. Protective effects of carnosine and homocarnosine on ferritin and hydrogen peroxide-mediated DNA damage. *BMB Reports* **43**:10, 683-687. [[CrossRef](#)]
18. Jung-Hoon Kang. 2010. Oxidative Damage of DNA Induced by Ferritin and Hydrogen Peroxide. *Bulletin of the Korean Chemical Society* **31**:10, 2873-2876. [[CrossRef](#)]
19. Aaron K. Holley, Yong Xu, Daret K. St. Clair, William H. St. Clair. 2010. RelB regulates manganese superoxide dismutase gene and resistance to ionizing radiation of prostate cancer cells. *Annals of the New York Academy of Sciences* **1201**:1, 129-136. [[CrossRef](#)]

20. Aaron K. Holley, Sanjit Kumar Dhar, Yong Xu, Daret K. St. Clair. 2010. Manganese superoxide dismutase: beyond life and death. *Amino Acids* . [[CrossRef](#)]
21. Hung-Hwan Yoon, Myeong-Seon Lee, Jung-Hoon Kang. 2010. Reaction of ferritin with hydrogen peroxide induces lipid peroxidation. *BMB Reports* **43**:3, 219-224. [[CrossRef](#)]
22. Anupam Bishayee, Themis Politis, Altaf S. Darvesh. 2010. Resveratrol in the chemoprevention and treatment of hepatocellular carcinoma. *Cancer Treatment Reviews* **36**:1, 43-53. [[CrossRef](#)]
23. Ehab H. Sarsour , Maneesh G. Kumar , Leena Chaudhuri , Amanda L. Kalen , Prabhat C. Goswami . 2009. Redox Control of the Cell Cycle in Health and Disease. *Antioxidants & Redox Signaling* **11**:12, 2985-3011. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
24. Georg T. Wondrak . 2009. Redox-Directed Cancer Therapeutics: Molecular Mechanisms and Opportunities. *Antioxidants & Redox Signaling* **11**:12, 3013-3069. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
25. P. R. Deepa, V. Nalini, K. Mallikarjuna, S Vandhana, S Krishnakumar. 2009. Oxidative Stress in Retinoblastoma: Correlations with Clinicopathologic Features and Tumor Invasiveness. *Current Eye Research* **34**:12, 1011-1018. [[CrossRef](#)]
26. Bridget Walsh, Amanda Pearl, Sarah Suchy, John Tartaglio, Kristin Visco, Shelley A. Phelan. 2009. Overexpression of Prdx6 and resistance to peroxide-induced death in Hepa1-6 cells: Prdx suppression increases apoptosis. *Redox Report* **14**:6, 275-284. [[CrossRef](#)]
27. Giovambattista Pani , Elisa Giannoni , Tommaso Galeotti , Paola Chiarugi . 2009. Redox-Based Escape Mechanism from Death: The Cancer Lesson. *Antioxidants & Redox Signaling* **11**:11, 2791-2806. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
28. Michael J. Hitchler, Frederick E. Domann. 2009. Metabolic defects provide a spark for the epigenetic switch in cancer. *Free Radical Biology and Medicine* **47**:2, 115-127. [[CrossRef](#)]
29. Alba Minelli, Ilaria Bellezza, Carmela Conte, Zoran Culig. 2009. Oxidative stress-related aging: A role for prostate cancer?. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer* **1795**:2, 83-91. [[CrossRef](#)]
30. Aaron K Holley, Daret K St Clair. 2009. Watching the watcher: regulation of p53 by mitochondria. *Future Oncology* **5**:1, 117-130. [[CrossRef](#)]
31. Joydeb Kumar Kundu, Hye-Kyung Na, Young-Joon Surh Intracellular Signaling Molecules as Targets of Selected Dietary Chemopreventive Agents **20083666**. . [[CrossRef](#)]
32. K Lei, D M Townsend, K D Tew. 2008. Protein cysteine sulfinic acid reductase (sulfiredoxin) as a regulator of cell proliferation and drug response. *Oncogene* **27**:36, 4877-4887. [[CrossRef](#)]
33. Dimitrios Galaris, Vasiliki Skiada, Alexandra Barbouti. 2008. Redox signaling and cancer: The role of "labile" iron. *Cancer Letters* **266**:1, 21-29. [[CrossRef](#)]
34. Jianli Chen, Mahesha Adikari, Rajash Pallai, Hemant K. Parekh, Henry Simpkins. 2008. Dihydrodiol dehydrogenases regulate the generation of reactive oxygen species and the development of cisplatin resistance in human ovarian carcinoma cells. *Cancer Chemotherapy and Pharmacology* **61**:6, 979-987. [[CrossRef](#)]
35. Ivan Bogeski, Valentin Mir#eski, Markus Hoth. 2008. Probing the redox activity of T-lymphocytes deposited at electrode surfaces with voltammetric methods. *Clinical Chemistry and Laboratory Medicine* **46**:2, 197-203. [[CrossRef](#)]
36. Elizabeth Anne Hillard, Fabiane Caxico de Abreu, Danielle Cristhina Melo Ferreira, Gérard Jaouen, Marília Oliveira Fonseca Goulart, Christian Amatore. 2008. Electrochemical parameters and techniques in drug development, with an emphasis on quinones and related compounds. *Chemical Communications* :23, 2612. [[CrossRef](#)]
37. Marco Giorgio, Mirella Trinei, Enrica Migliaccio, Pier Giuseppe Pelicci. 2007. Hydrogen peroxide: a metabolic by-product or a common mediator of ageing signals?. *Nature Reviews Molecular Cell Biology* **8**:9, 722-728. [[CrossRef](#)]
38. Ana García-Navarro, Cristina González-Puga, Germaine Escames, Luis C. López, Ana López, Manuel López-Cantarero, Encarnación Camacho, Antonio Espinosa, Miguel Angel Gallo, Darío Acuña-Castroviejo. 2007. Cellular mechanisms involved in the melatonin inhibition of HT-29 human colon cancer cell proliferation in culture. *Journal of Pineal Research* **43**:2, 195-205. [[CrossRef](#)]
39. Heung-Jai Park, Seong-Wook Jeong, Jong-Myoung Kim, Won-Gun An. 2007. Detection of Methyl ethyl ketone in the Ambient Air of Industrial Area in Gimhae City and Its Effect on the Generation of Reactive Oxygen Species. *Journal of the Environmental Sciences* **16**:8, 995-999. [[CrossRef](#)]